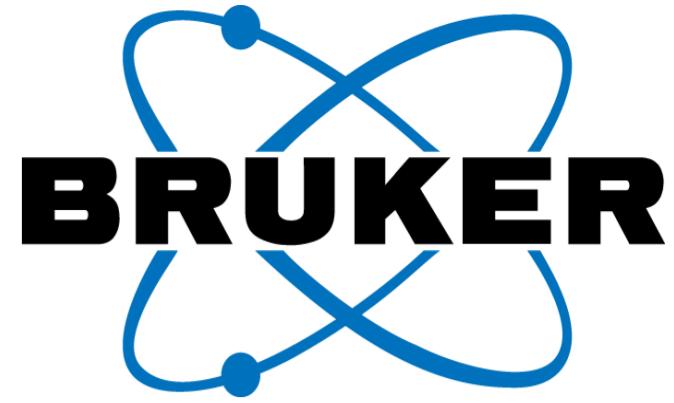


# 4D-Lipidomics™ based automated annotation of MALDI Imaging data using a dedicated bioinformatics pipeline

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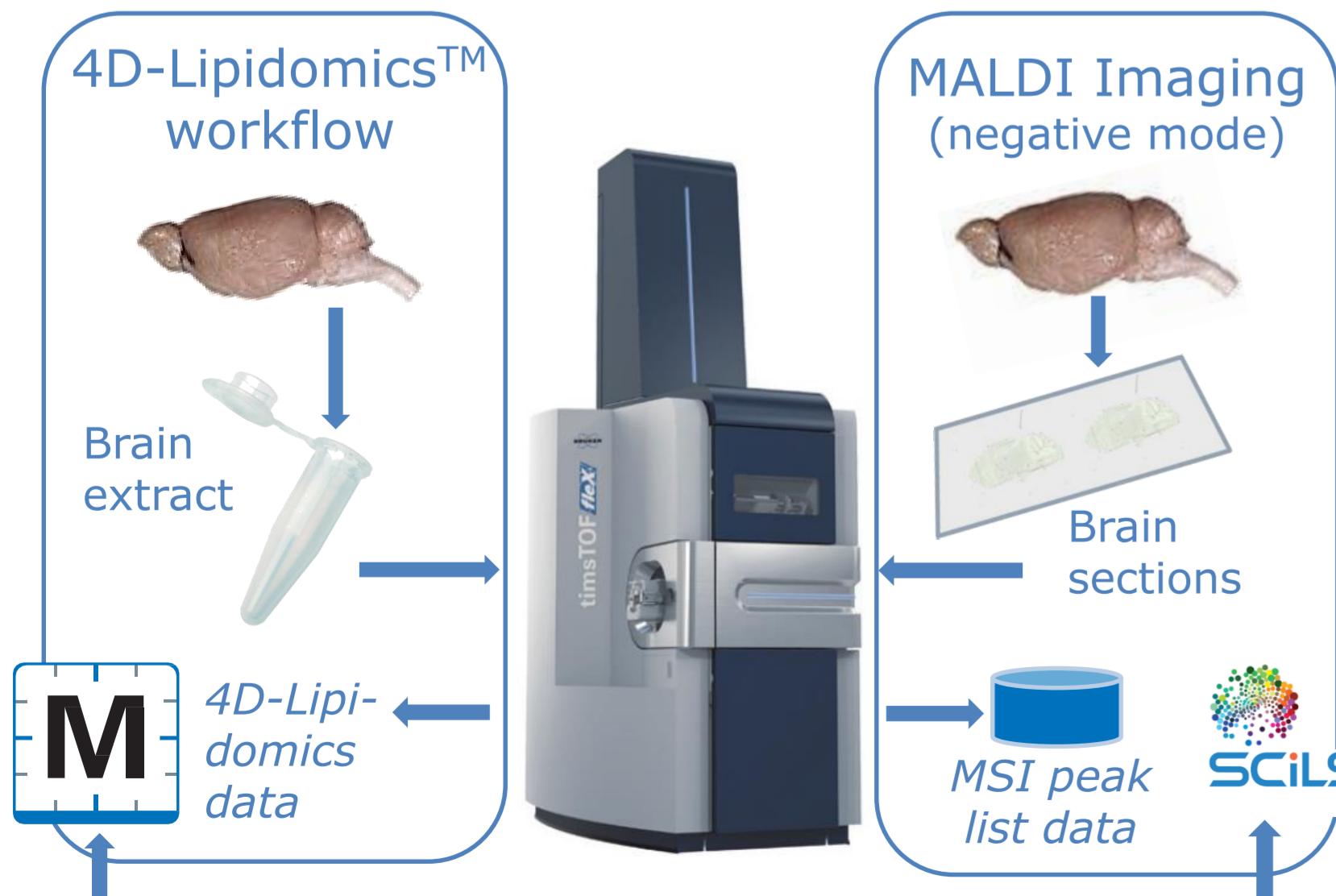
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## Introduction

The complexity of lipid samples makes their analysis directly from tissue sections extremely challenging. Ion mobility helps in this respect by introducing an additional analytical dimension that can separate isomeric and isobaric molecules. Moreover, annotation of lipids from MALDI Imaging data is often hampered by the limited ability to perform large-scale MS/MS experiments directly from tissue. Here we demonstrate a workflow combining 4D-Lipidomics™ and MALDI Imaging data obtained from just one instrument, timsTOF fleX, in conjunction with a bioinformatics pipeline for automatic lipid annotation.

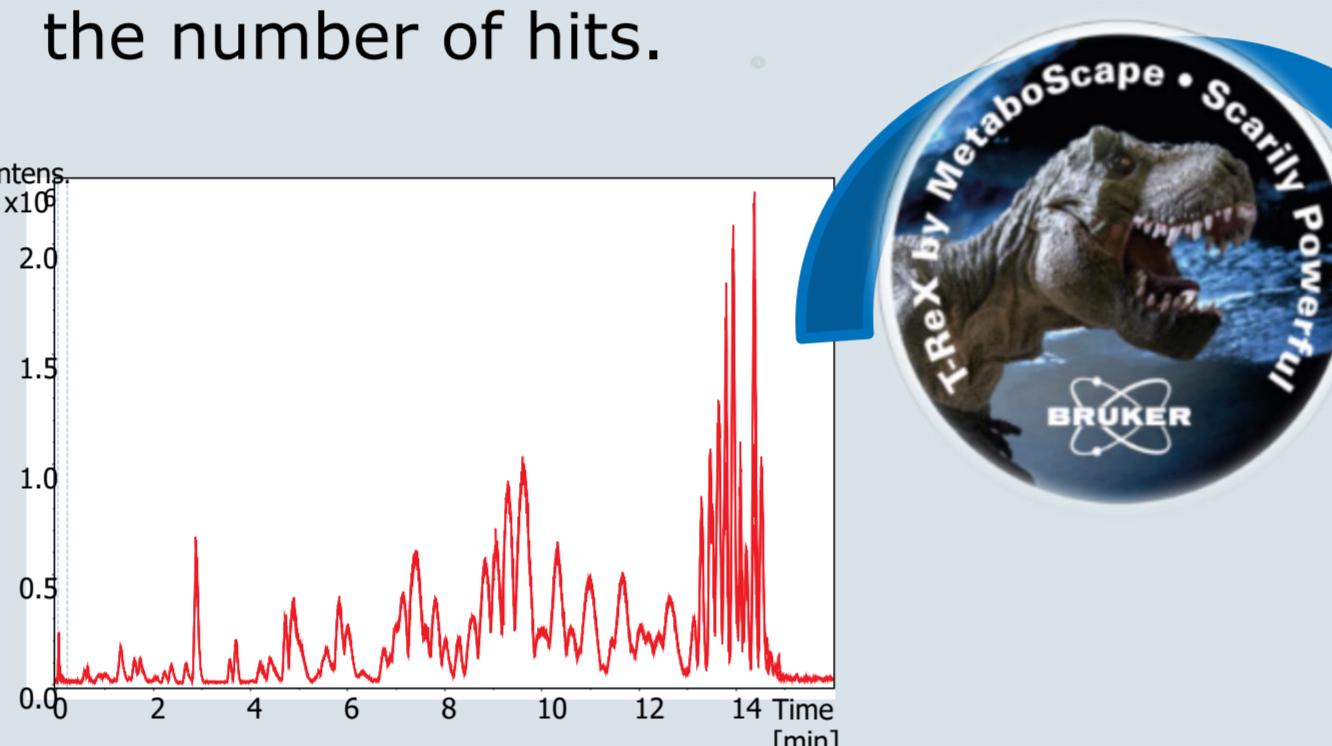
## Methods



## Results

### 4D-Lipidomics results

Apply five quality criteria on PASEF empowered 4D-Lipidomics data for confident annotations. Merge positive and negative mode data to increase the number of hits.



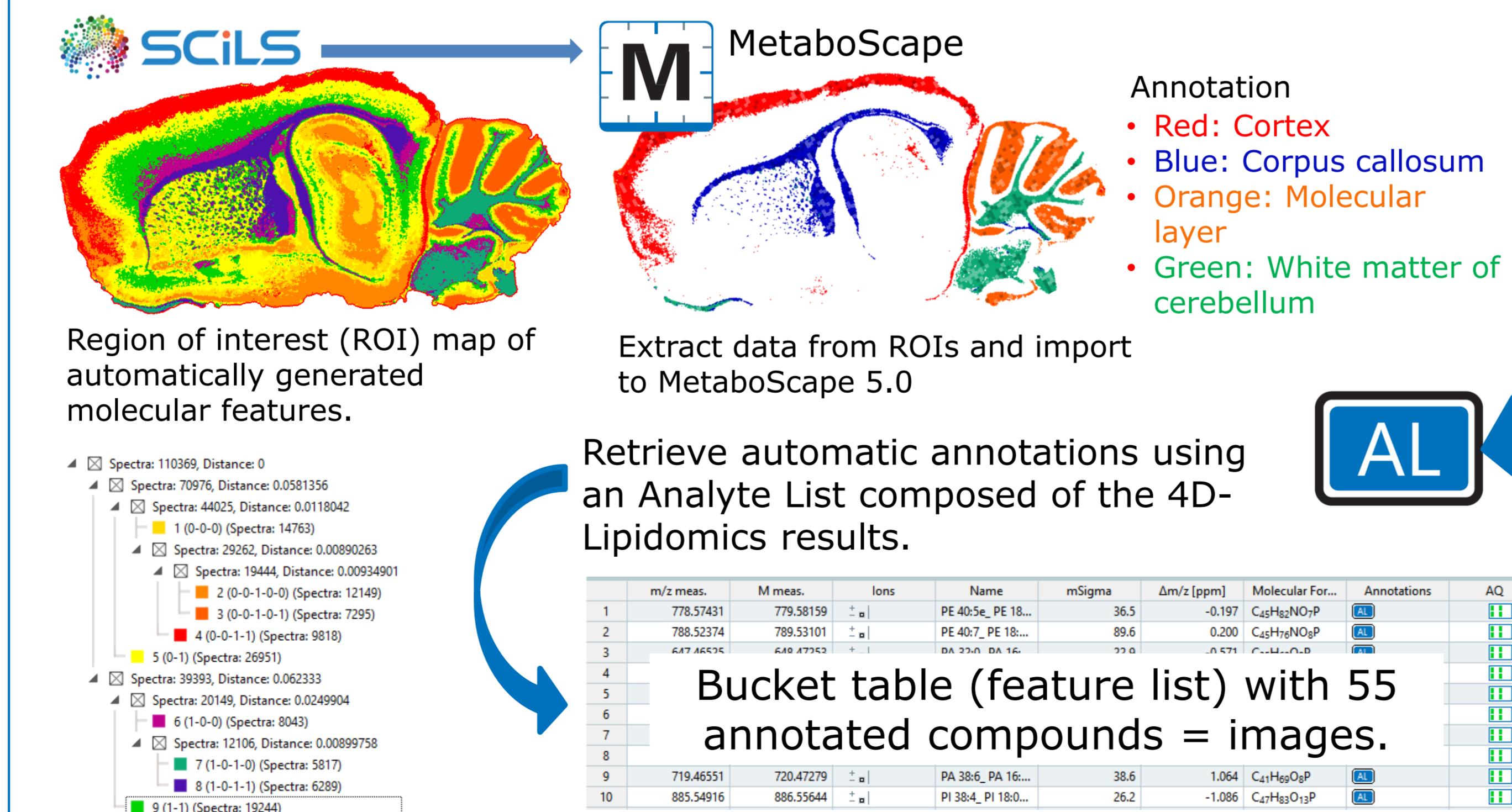
### AQ score



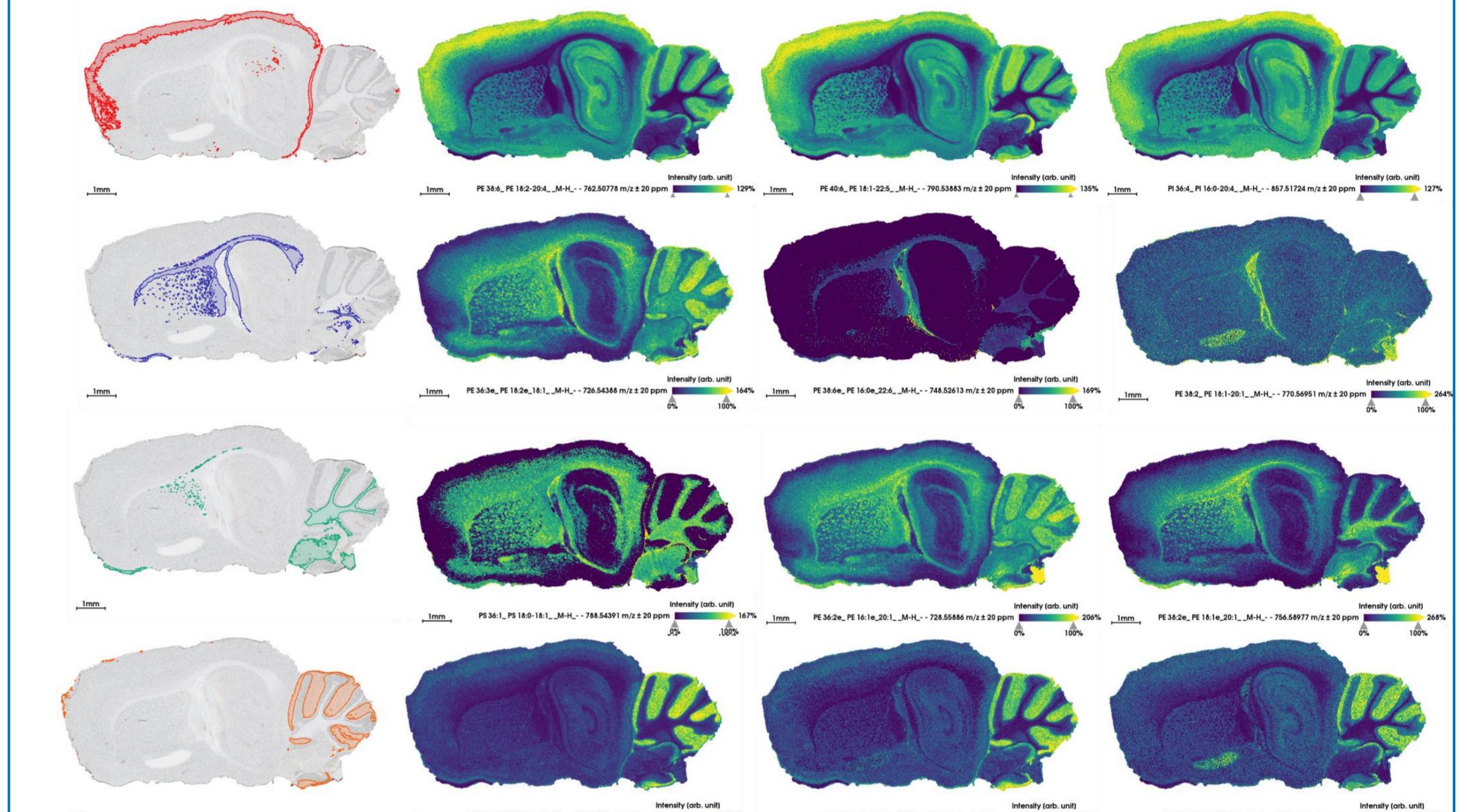
- A Precursor mass accuracy
- B Retention time
- C Isotopic pattern
- D MS/MS fragment spectra
- E CCS values

	Data	Buckets	Buckets with MS/MS	Annotations based on LipidBlast Search
Pos. Ionization	7107	2984	811	
Neg. Ionization	2915	1535	546	
Pos / Neg merged	9797	4353	1232	

### MALDI Imaging results.

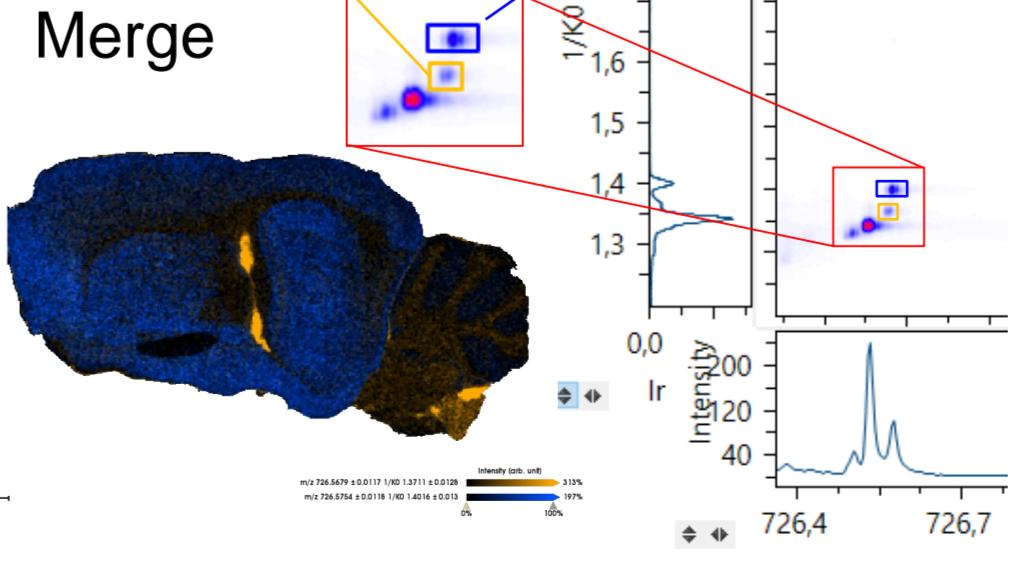
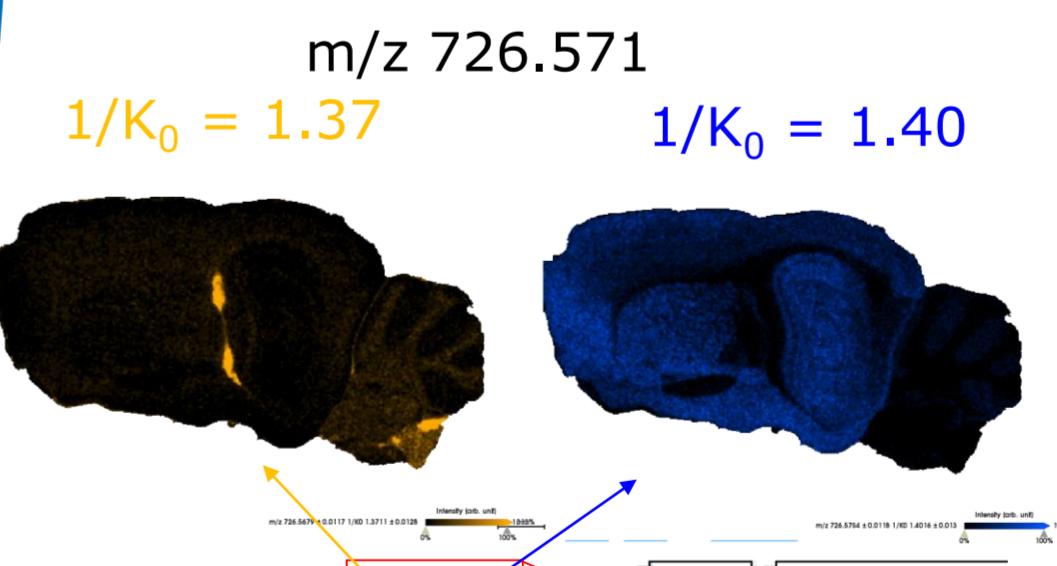


### Correlate molecular information with spatial information.



### 4D-Molecular Imaging

Separate isobaric or isomeric compounds using trapped ion mobility spectrometry (TIMS).



timsTOF fleX

### Conclusions

- Highly confident annotations of PASEF empowered 4D-Lipidomics™ data were retrieved using five quality scoring criteria.
- SpatialOMx® workflow for automatic annotation of MALDI Imaging data using SCiLS Lab and MetaboScape presented.
- 4D-Molecular Imaging applying TIMS reveals distributions of isobaric compounds without having hundreds of thousands of resolving power.