



PaSER 2022

- Search Engine in Real-time

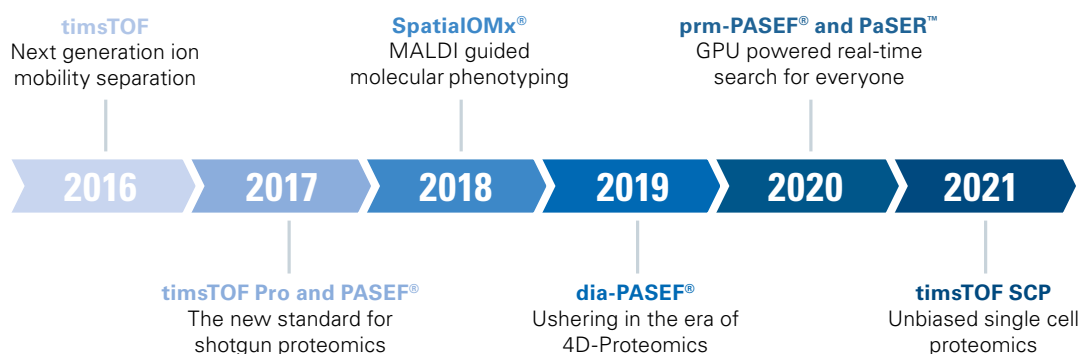
PaSER 2022

GPU powered Parallel Search Engine in Real-time



From data acquisition to data analysis your lab's productivity is critical. timsTOF platform enables you to run more experiments with greater information content than ever before, but more experiments mean more data to analyze. By introducing real-time search capabilities with PaSER™ you can now break the data analysis bottleneck. PaSER™ brings you Run & Done with smarts, where at the end of every data acquisition you have results in hand, maximizing your labs efficiency and making the most informed acquisition decision.

Proteomics Innovations by Bruker

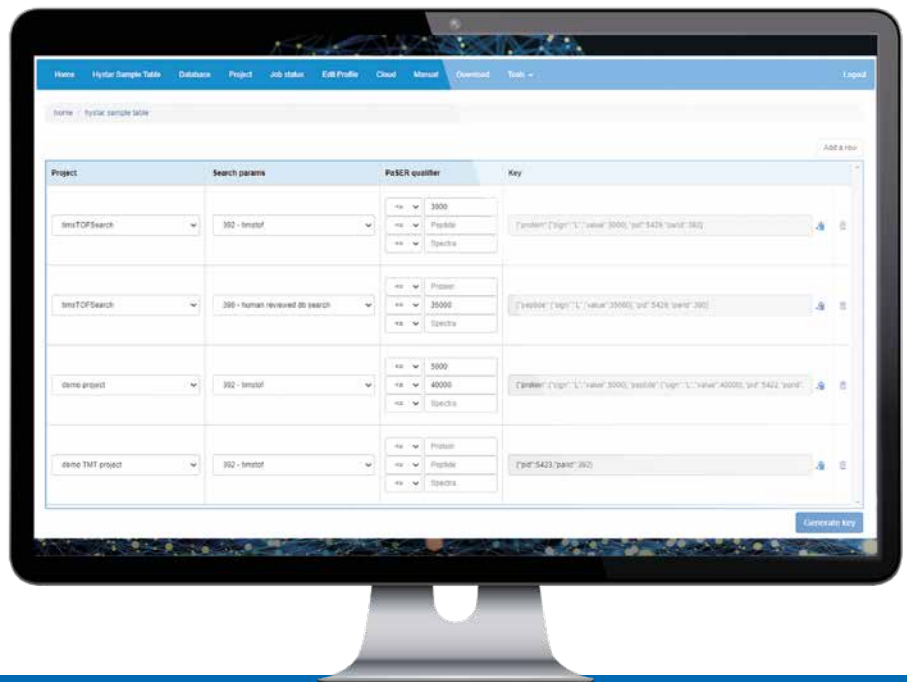


Bruker innovations in recent years have transformed the proteomics landscape, starting with the timsTOF Pro which ushered in the era of 4D-Proteomics™. 4D-Proteomics enabled by PASEF® provides MS/MS acquisition at >100 Hz, making high-throughput measurements using **short gradients** possible while maintaining **deep proteome coverage** or high protein depth measurements in **less time**. This gives you the most information rich datasets for your samples in the least amount of time.

The big picture that Bruker is delivering with timsTOF and PaSER

Sequencing of the human genome was thought to have revolutionized the application of not only genomics but also proteomics, because the description for each coded gene could be predicted. Formerly, proteomics struggled to achieve expectations for reasons including lack in proteomic depth of coverage, quantitative capabilities, and throughput. The capabilities of modern mass spectrometers, most notably the timsTOF Pro 2, have changed this outlook. Instead of analyzing tens of samples per day at maximal sample loads (>1 µg) with ineffective sequencing speeds (<40 Hz), the timsTOF Pro has allowed hundreds of samples with minimal sample load (<200 ng) at sequencing speeds exceeding 100 Hz. All of this with uncompromised proteomic depth. This combination has changed the way proteom-

ics can be done, but it also unleashes a new constraint, which is data analysis. Formerly having to process tens of samples in a day was an easy task, but modern researchers with timsTOF technology must process hundreds of samples of information dense content. For most pipelines this creates a bottleneck at the data analysis step, for which we have specifically tried to innovate. Bruker has done so by introducing real-time database search capabilities called PaSER (Parallel Search Engine in Real-time) which removes the data analysis hurdle. Using PaSER, as soon as a LC-MS run is completed, results are in hand or Run & Done.



Rest easy, knowing PaSER is doing the work for you:

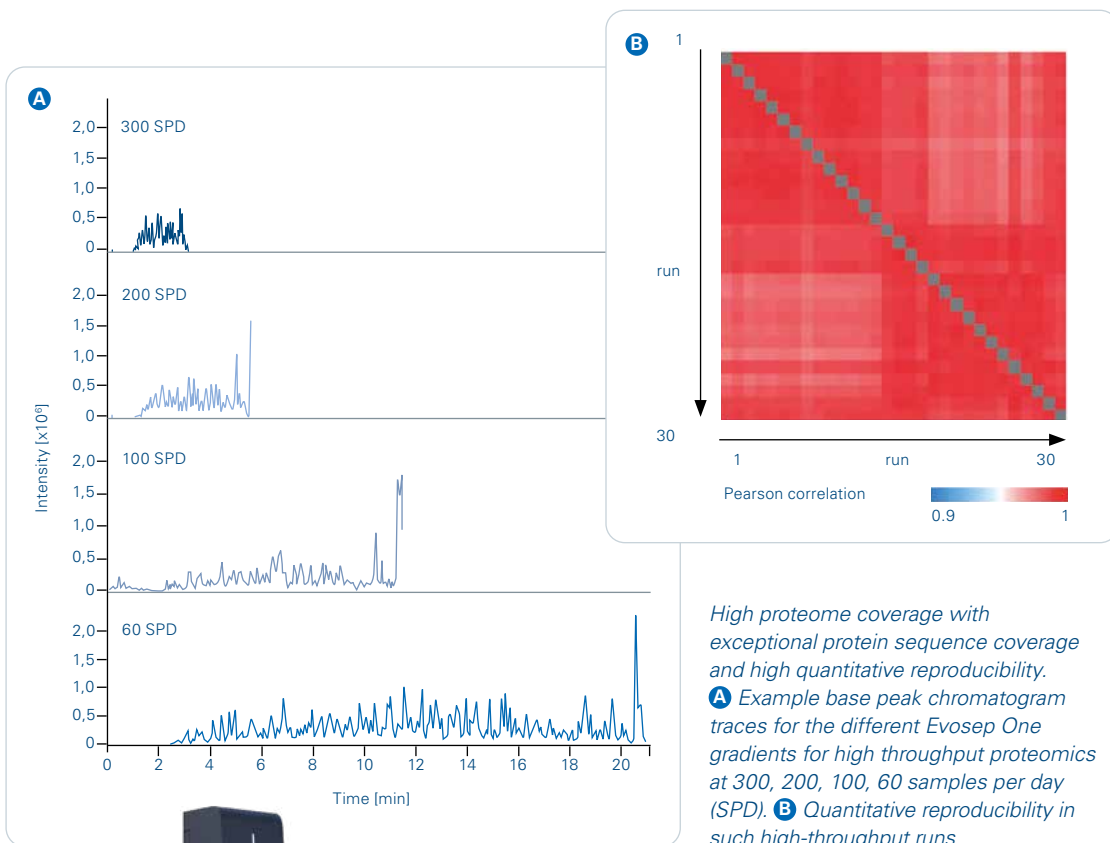
Several groups have envisioned the utility of a system that makes use of prior knowledge to make decisions 'on-the fly'. PaSER makes this a reality. PaSER is a complete hardware and software solution that brings intelligent acquisition to the lab and to any current or new timsTOF instrument.

High-throughput 4D-Proteomics

by coupling short gradients to PASEF technology

PASEF delivers the synergistic benefits of sensitivity and speed. This combined with short gradients results in unprecedented proteome coverage without compromise in protein sequence coverage. The sheer speed of PASEF

(120 Hz) has pushed the formerly accepted “norm” of two hour gradients to less than 30 minutes. Now proteomes can be measured at a pace of 60, 100, 200 and 300 sample measurements in a single day.



High proteome coverage with exceptional protein sequence coverage and high quantitative reproducibility.

A Example base peak chromatogram traces for the different Evosep One gradients for high throughput proteomics at 300, 200, 100, 60 samples per day (SPD). **B** Quantitative reproducibility in such high-throughput runs.



This paradigm shift in gradient lengths used for proteomics naturally leads to measuring many more samples. This transfers the capability of large-scale proteomic systems biology studies and large cohort clinical research applications to any lab equipped with a timsTOF mass spectrometer.

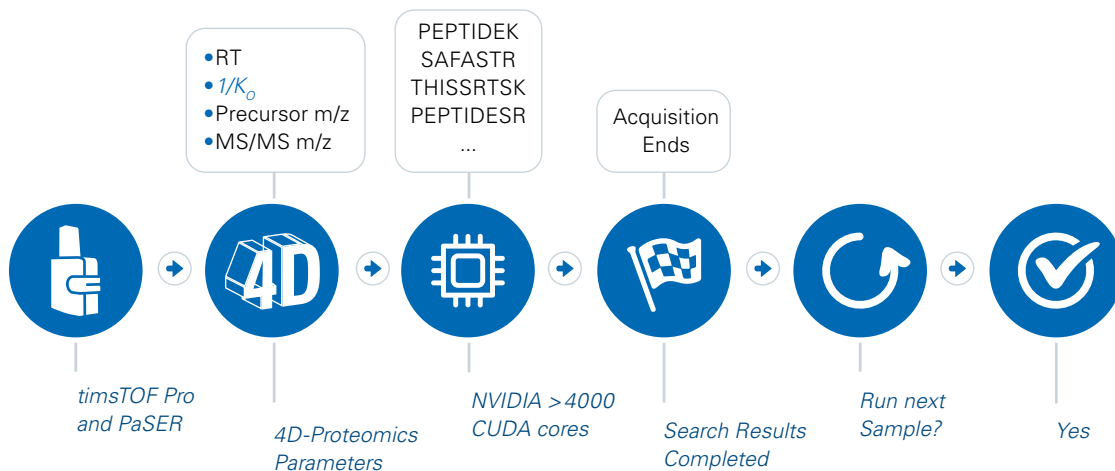
Introducing PaSER Run & Done

Uncompromised data processing on the fly

Traditionally data acquisition has been the bottleneck for large scale proteomics. This has also remained one of the limitations in leveraging mass spectrometry within the clinic. PASEF and short gradients have facilitated these large-scale studies in any lab but have pushed the bottleneck downstream to data processing. The timsTOF Pro 2 has made the acquisition of thousands of samples in a modest period of time reality. Thus, it is now crucial to have real-time results particularly in terms of identification to monitor the quality of datasets and intervene in a timely manner when required. This is made possible by Parallel Search Engine in Realtime (PaSER).

PaSER 2022 is a complete hardware and software solution that has two focuses: Speed and Smarts.

PaSER is powered by a GPU optimized search algorithm and necessary hardware to communicate to the instrument's acquisition PC to identify peptide sequences from the MS/MS spectra as they 'elute' from the acquisition PC. These results are then appended, and actions can be triggered based on the cumulative identification results at the end of the sample measurement.



The PaSER workflow – simple, fast and smart: Users interface with the acquisition PC to setup an acquisition table. In addition to standard parameters, such as injection volume and LC-MS methods, users would also specify a PaSER method and qualifiers for each injection. As data is acquired on the acquisition PC, the data is simultaneously streamed to the PaSER box. This 4D data is processed by PaSER in real-time providing results as soon as acquisition ends enabling Run & Done. These results are utilized to verify if the user defined qualifier was met and if acquisition should continue.

No more worrying about injecting those precious samples. No more worrying of wasted instrument time. No more worrying about those samples with expensive reagents and sample prep, only to inject on a poor performing system. No more waiting for the QC runs to determine system suitability. No more waiting for search results during method development.

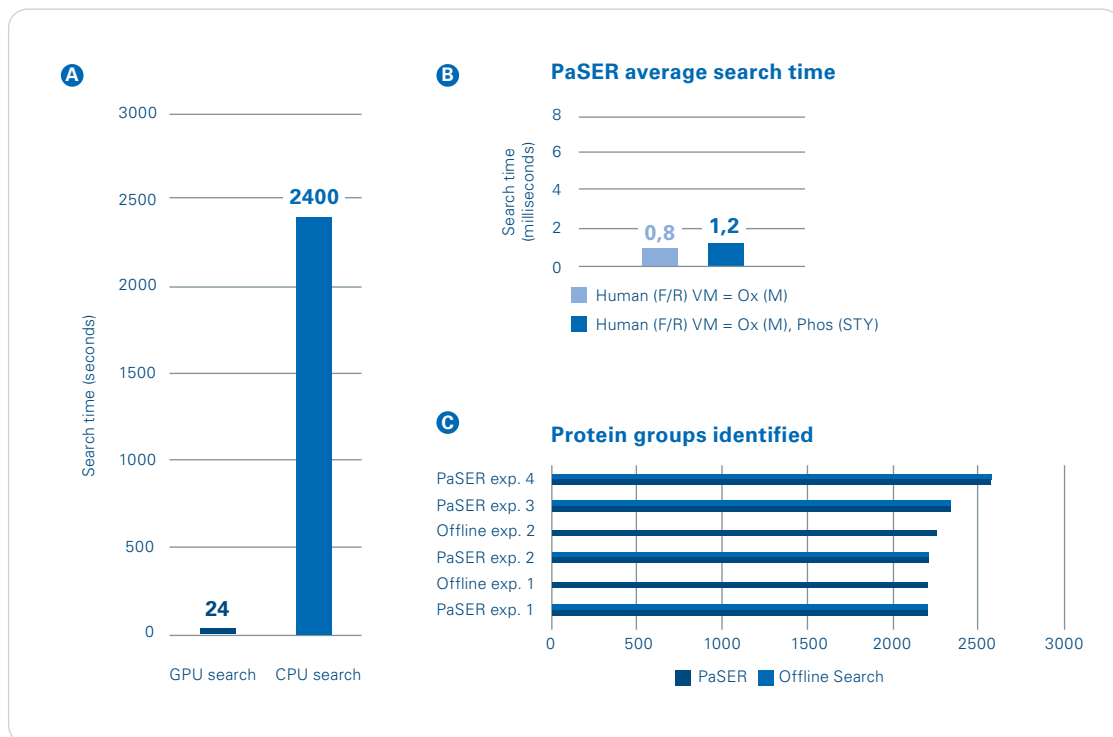


- ✓ Precious samples
- ✓ Instrument time
- ✓ Analysis time

Full-fledged database search in less than 5 ms

PaSER provides consistent results obtained from the real-time search, identical to offline search of the same data. Since the database search algorithm is run on the GPU, the search times

are negligible compared to CPU based searches. In addition to real-time search the opportunity to search a larger space for specific applications, including peptidome studies, becomes reality.

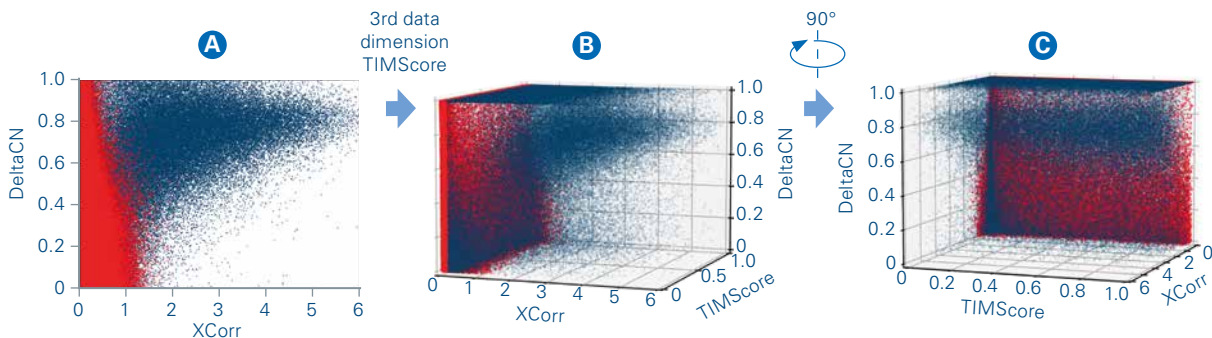


Blazing search speed. **A** A single injection of a human cell lysate analyzed against the forward and reverse sequences of the Uniprot Human fasta file with up to 6 PTMs using either ProLuCID or ProLuCID-GPU search algorithm. **B** PaSER can process MS/MS spectra with an average search time ~ 1 ms. **C** PaSER search results are nearly identical whether done in a real-time manner or offline.

TIMScore – Machine Learning and CCS exploited for better FDR

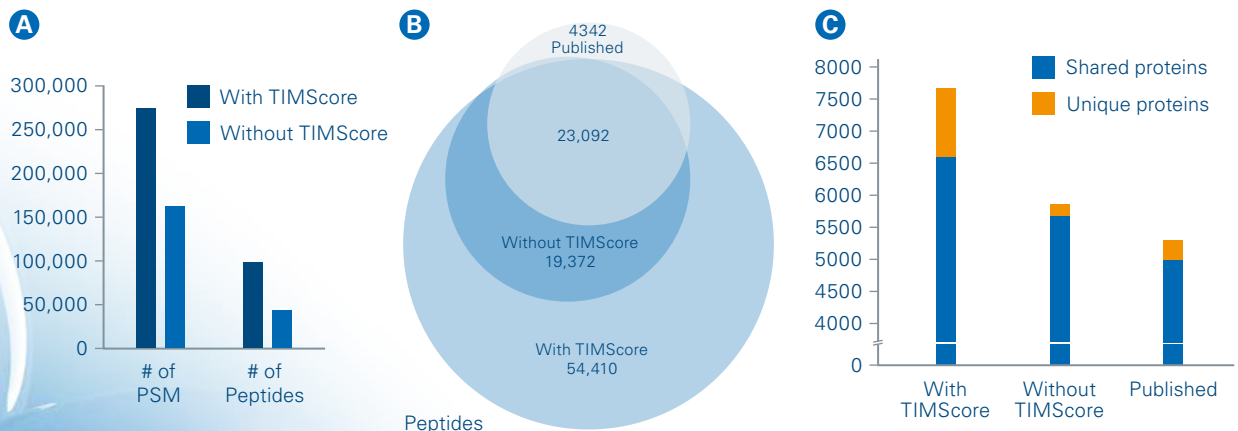
The first database search algorithm, developed in the Yates lab circa 1994, revolutionized proteomics and utilized the cross-correlation (XCorr) scoring function for identifying peptide MS/MS spectrum. XCorr is still widely utilized or adapted in many popular algorithms including Comet, Crux Tide and of course ProLuCID. TIMScore extends that concept to the mobility dimension. TIMScore is the measure of the correlation

between the experimentally measured CCS for a peptide versus predicted CCS of that peptide. To predict the CCS of peptides, deep learning was utilized to build a transformer model, which utilizes self-attention to differentially weight the significance of each part of the input data, providing a fast and robust model of CCS values when provided with the amino acid sequences.



Utilizing TIMScore is easy

The PaSER search algorithm is run as normal, and the search algorithm automatically compares the predicted and measured CCS values and calculates TIMScore for the top 5 peptide candidates for each spectra. The true benefit of TIMScore can be realized during the peptide-validation and False Discovery Rate (FDR) estimation steps of the proteomics pipeline. In a non CCS-enabled algorithm, only two dimensions can be utilized to estimate the FDR rate, and so a discriminate line is fit to a 1% error (Panel A) to distinguish forward and reverse peptide candidates. With TIMScore, and the extra CCS dimension, the peptide-candidates can be vectorized in 3 dimensions (Panel B) allowing a discriminate contoured plane to be applied to achieve the same 1% error. Applying a discriminate plane provides increased accuracy and precision, helping to validate formerly poorly scoring PSMs in the standard two dimensions. Thus, the key effect of TIMScore is derived from the additional dimension of CCS it provides in assigning true positives from decoy peptide sequences as shown above.



TIMScore increases proteins, peptides, PSMs and sequence coverage

TIMScore when applied to the published data set from the laboratory of Prof. Yasushi Ishihama titled "Effect of Phosphorylation on the Collision Cross Sections of Peptide Ions in Ion Mobility Spectrometry" adds more than 110,000 PSMs and doubles the number of peptides observed from 42,930 to 98,949. The > 98,000 peptides observed for this dataset is a 3.5 times increase in what was reported in the published data and shows good overlap with regards to protein groups and peptides identified (Panel B and C). TIMScore significantly boosts protein sequence coverage which helps provide quantitative accuracy and more expansive libraries for

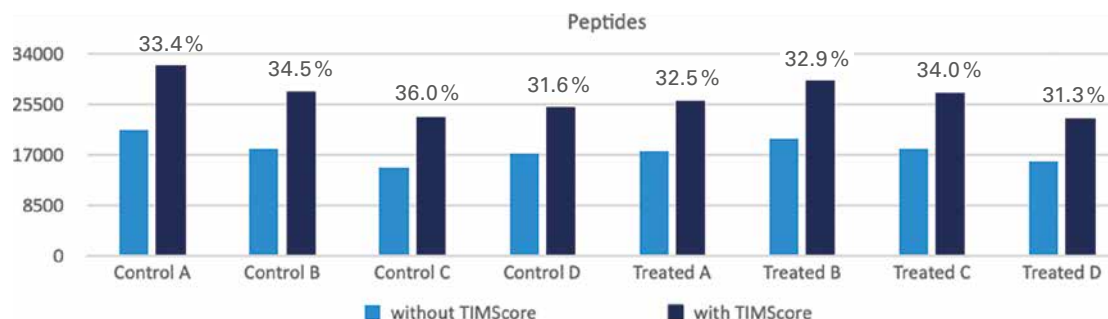
label free quantitation (LFQ), data independent acquisition (DIA) and parallel reaction monitoring (PRM) experiments.

Similarly, when TIMScore is applied to an unpublished AP-MS study from the laboratory of Prof. Stanley Stevens Jr. the number of peptides identified increases by > 30 % in all 4 control and all 4 treated samples (see graph below). Without TIMScore 24,664 phosphopeptides were identified across all 8 samples, which was doubled to 49,423 phosphopeptides with TIMScore. Furthermore, 19,159 of these phosphosites were uniquely identified in the treated samples and not the control samples.

TIMScore increases phosphopeptide identification and phosphosite localization

More identifications are meaningless if the site of the PTM cannot also be determined. Luci-PHO, a tool to assess the number and accuracy of PTM sites via false localization rate (FLR) [Fermin 2013] was used to determine the percent confidence interval of correct assignment of phosphorylated peptides. Consistent with

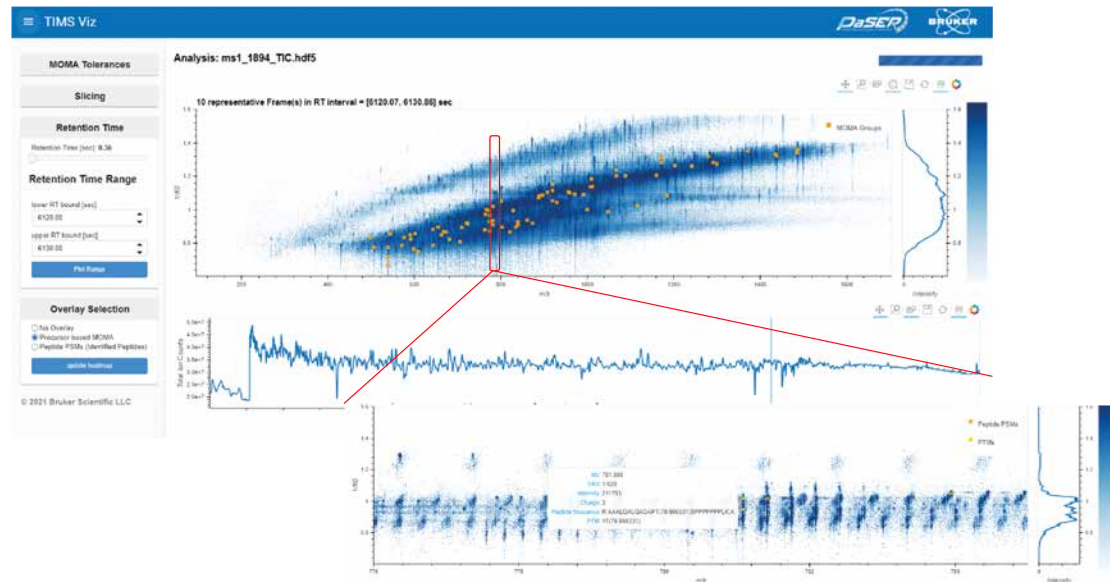
the increased sensitivity TIMScore provides, it improved the number of localized phosphopeptide PSMs at the 5 % FLR by 10,854 or 57 %. These percent improvements using TIMScore held consistent (57-62 %) across both PSMs and peptides at both the 5 % and 1 % FLR.



"CCS values and 4D-Proteomics™ have become indispensable in our application of mass spectrometry-based proteomics. TIMScore shows promise to transform how DDA searches are performed, identifying many more meaningful peptides and proteins allowing us to go deeper into the proteome using CCS."

Professor Stanley Stevens Jr. at the University of South Florida

TIMS Viz – Effortlessly identify isobaric masses resolved by mobility



TIMS Viz provides an effortless way to visualize timsTOF data. Since its launch in 2017 the timsTOF has provided mobility measurements for all peptide, lipid and small molecule analytes which have proven to be reproducible across instruments and laboratories. Until the introduction of TIMS Viz multi-dimensional visualization was difficult. Using TIMS Viz the data acquired on PaSER can be displayed, parsed, and interrogated allowing you to finally see co-eluting, isomeric or near-isomeric ions which collapse onto a single spectrum and remain hidden without mobility separation. TIMS Viz can also overlay the real-time search results from a PaSER run, presenting identified peptide sequences by retention time, m/z and mobility values.

One of the most powerful features within TIMS Viz is the MOMA viewer. Complex protein digests contain hundreds of thousands and even millions of peptides. In running them by LCMS

it is common that ions of a similar m/z co-elute in time. When this happens a traditional mass spectrometer is unable to deconvolute these ions as two species, resulting in co-fragmentated MSMS spectra. This is where the power of TIMS shines, when those two co-eluting ions have even the smallest difference in their CCS then they will be represented by different mobility (CCS) values and interrogated as separate species. This advantageous feature is called mobility offset mass aligned or MOMA. The TIMS Viz example above is a 10 second retention time extraction where within this time window many MOMA groups (yellow dots) have been detected. The zoomed window displays a phosphopeptide positional isomer, where both the serine and threonine sites have been positively identified by MS/MS, co-eluting in time and resolved only by their CCS which are different.

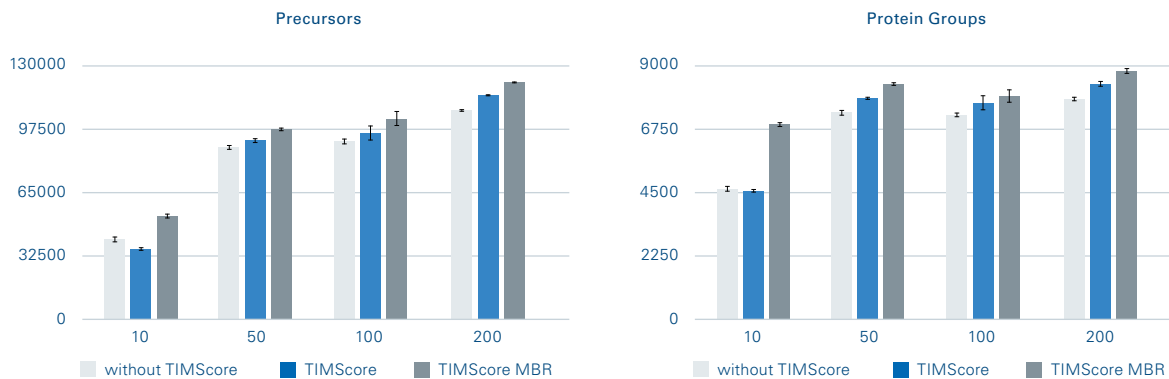
dia-PASEF workflows with TIMS DIA-NN

PaSER 2022 is capable of processing dia-PASEF workflows utilizing a customized version of the popular DIA-NN (DIA by Neural Networks) software from the Lilley, Rasler and Demichev labs. TIMS DIA-NN enables reliable, robust, and quantitatively accurate large-scale experiments. TIMS DIA-NN provides stringent statistical control, flexible modelling (AE) of data and automatic parameter optimization.

PaSER utilizes the same stream mechanism for dia-PASEF workflows, streaming in real-time MS and DIA frames from the acquisition PC to the

PaSER box. These dia-PASEF data are processed and stored on the PaSER box. At the end of the acquisition a spectral library search is triggered, and the results recorded. TIMScore powered DDA search results can be easily utilized to build spectral libraries or spectral libraries can be imported from other popular tools. At the end of the project the users can trigger match-between-runs analysis of the full project (or selected acquisitions) to view a quantitative profile across the project. This provides users an integrated environment for PASEF and dia-PASEF data analysis.

Nearly 9000 protein groups identified from 200 ng of K562 lysate in 35 min gradient (~30 samples per day) with TIMS DIA-NN and a TIMScore spectral library.



"Our ongoing collaboration with Bruker to tailor DIA-NN to a streamlined processing tool for dia-PASEF data with a CCS focus has been really rewarding. It simplifies and accelerates identifying and quantifying thousands of proteins in even very short gradients. We are pleased that within our close collaboration with Bruker, the vendor-integrated version of DIA-NN called TIMS DIA-NN now becomes part of the PaSER bioinformatics platform."

Prof. Dr. Markus Rasler, Einstein Professor of Biochemistry at Charité, Berlin, Germany

PaSER Data Viewer

Examine your data however you like

PaSER is complimented with a full featured, LIMS based analysis platform and data viewer. All real-time search results are stored on the PaSER box. Users may examine the results in as much detail as they desire. Start with a simple overview of the results, dig deeper into a specific protein, or visualize an individual MS/MS spectrum. Users can

also re-search past acquisitions, perhaps with an updated fasta file, a new PTM or simply apply a new FDR threshold. Search results from multiple samples can be combined and comparative analysis can be conducted. PTM localization scores and their MS/MS spectra can be visualized. The PaSER Data Viewer affords you the capability to decide.

Experimental summary

ID	Search Results	Database	PSM Score	Name	Misc. quality check	Searched Scans/Total Scans	Scan Ratio	data	peptide count	proteins	peptide	proteins	peptide	spectrum
155700	Result View	prot	prot	hms2TOF_Human_msc_4786	View	281853 / 282295	97.0 %	2020-09-08	1	5507	74100	9.98	0.15	0.1

Protein ID

protein	accession	Gene Name	seq coverage	seq count	NDC Count	length	MW	NAF	empsi	description
view	ADA024R801	NUOT4B	58.1	9	24	181	20434.0	4.0883944E-4	2.8994195	Diphosphonitrosyl polyphosphate phosphohydrolase NUOT4B OS=Homo sapiens OX=9908 GN=NUOT4B PE=3 SV=1
view	ADA084J2D5	GATD3B	34.7	9	25	268	28142.0	2.8762428E-4	1.22331	Glutamine amidotransferase-like class 1 domain-containing protein 3B, mitochondrial OS=Homo sapiens OX=9908 GN=GATD3B PE=1 SV=1
		GATD3A	34.7			268	28170.0	2.8762428E-4	1.22331	Glutamine amidotransferase-like class 1 domain-containing protein 3A, mitochondrial OS=Homo sapiens OX=9908 GN=GATD3A PE=1 SV=1
view	ADA0JWYVL9	TEX13C	4.1	2	2	993	108809.0	6.219136E-8	0.0990582	Putative testis-expressed protein 13C OS=Homo sapiens OX=9906 GN=TEX13C PE=5 SV=1
view	ADA0U1RRL7	MMP2A09	74.8	2	6	71	7679.0	2.8956328E-4	4.5718575	Protein MMP2A09 OS=Homo sapiens OX=9909 GN=MMP2A09 PE=1 SV=1

Sequence coverage

Protein

accession: ADA024R801 sequence coverage (%) 58.56% description: Diphosphonitrosyl polyphosphate phosphohydrolase NUOT4B OS=Homo sapiens OX=9908 GN=NUOT4B PE=3 SV=1

Export options: CSV | Excel | AML | PDF

ADA024R801

0 50 100 150 200 250 300 350 400 450 500 550 600 650 700 750 800 850 900 950 1000 1050 1100 1150 1200 1250 1300 1350 1400 1450 1500 1550 1600 1650 1700 1750 1800 1850 1900 1950 2000 2050 2100 2150 2200 2250 2300 2350 2400 2450 2500 2550 2600 2650 2700 2750 2800 2850 2900 2950 3000 3050 3100 3150 3200 3250 3300 3350 3400 3450 3500 3550 3600 3650 3700 3750 3800 3850 3900 3950 4000 4050 4100 4150 4200 4250 4300 4350 4400 4450 4500 4550 4600 4650 4700 4750 4800 4850 4900 4950 5000 5050 5100 5150 5200 5250 5300 5350 5400 5450 5500 5550 5600 5650 5700 5750 5800 5850 5900 5950 6000 6050 6100 6150 6200 6250 6300 6350 6400 6450 6500 6550 6600 6650 6700 6750 6800 6850 6900 6950 7000 7050 7100 7150 7200 7250 7300 7350 7400 7450 7500 7550 7600 7650 7700 7750 7800 7850 7900 7950 8000 8050 8100 8150 8200 8250 8300 8350 8400 8450 8500 8550 8600 8650 8700 8750 8800 8850 8900 8950 9000 9050 9100 9150 9200 9250 9300 9350 9400 9450 9500 9550 9600 9650 9700 9750 9800 9850 9900 9950 10000

number of peptides overlapped number of spectra overlapped

MS2 visualization

Protein sequence: R ISQDEVLVLSR Y

Scan number: 104650

Calculated M+H: 1577.7653

Measured M+H: 1577.774

Charge state: 2

File name: 20191021_K562_200mg_30min_Site1-1_01_4796_nopt

search type: light

File: 20191021_K562_200mg_30min_Site1-1_01_4796_nopt, Scan: 104650, Precursor m/z: 789.1267, Charge: 2

SEQEDEVLYLSRR, MW= 1577.7653, m/z 789.1263

m/z	1	2	3	4
345.1405	1	1	1	1
376.1574	1	1	1	1
414.1421	1	1	1	1
449.1574	1	1	1	1
549.2108	1	1	1	1
575.1574	1	1	1	1
718.2216	1	1	1	1
750.2390	1	1	1	1
817.1219	1	1	1	1
900.4051	1	1	1	1
1043.4891	1	1	1	1
1162.2376	1	1	1	1
1229.5896	1	1	1	1
1314.5216	1	1	1	1
1403.4536	1	1	1	1
1386.4207	1	1	1	1

q1+	q1+	q1+	q1+	q1+	q1+
ac(slab)	m(z(measured))	m(z(calculated))	m(z(measured))	ppm	RetTime
132.6907	1332.9924	986.8428769140625	866.84983140625	1.3	3569.447
17.4069	887.4044	448.2071009322894	449.2054947260627	-2.8	1725.939
77.7653	1577.774	789.382650645313	789.396650257813	5.6	2887.312
81.1714	2541.1675	1271.089333398375	1271.087380234375	-1.5	4449.181
81.1714	2541.1650	847.72064890525	847.7207771814564	-2.2	4443.926
100.9581	1850.961	917.957992842709	917.959389587709	1.8	1132.834
100.776	3590.762	1200.56681256575	1200.592174208875	-3.9	5256.228
100.7996	3600.7678	1200.9247752083334	1200.9274607352084	2.2	5335.388

Examine your data and visualize everything. From an overview of the experiment down to individual MS/MS spectra. Compare across multiple samples or re-search acquired runs.

PaSER 2022

GPU powered Parallel Search Engine in Real-time



PaSER 2022 brings smart acquisition to your laboratory with a complete hardware and software solution.

The GPU based search algorithms in PaSER eliminates the searching bottleneck with real-time search.

TIMScore™ provides more confident peptide identifications.

TIMS Viz allows facile detection and visualization MOMA events.

dia-PASEF workflows are supported with an integrated version of **TIMS DIA-NN**.

Experience Run & Done. Additionally, PaSER can utilize the real-time search results to make "on-the fly decisions"



"Innovative software tools are a necessity to address unanswered biological questions with mass spectrometry. The trapped ion mobility functionality and the robustness of the timsTOF Pro offer unique bottom-up proteomics capabilities that can be effectively used to study many diseases."

Professor John Yates III, the Ernest W. Hahn Professor at The Scripps Research Institute in La Jolla, California

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