PREOMES

High-throughput and low-input Metabolomics/Lipidomics sample preparation – A new BeatBox[®] application

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SPOTLIGHT

- High-throughput approach that enables homogenization extraction from 1 to 96 samples.
- Fast, easy-to-use, and standardized sample preparation for r and lipid extraction from wide variety of tissue samples.
- Overall, excellent reproducibility determined by CV <20% analyt
- Facilitates the processing of very small amounts of tissue (w 1-5 mg), ideal for targeted MS approaches.
- Extraction of biologically relevant metabolites and lipids the second sec the various tissue types.



MATERIALS & METHODS

Tissue and sample preparation:

Approximately 5 mg wet weight tissue from mouse (cardiac muse intestine, liver, lung, muscle, and spleen). Using a 1.0 mm disposab pen (Integra Lifesciences), a single tissue punch was added to eac a BeatBox Tissue Kit 96x plate with 100 µl of a cold 80% methanol The tissue was homogenized for 10-minutes by using the E standard power settings.

After centrifugation (4,000g for 20-minutes at 4 °C) supernation collected and protein precipitate was discarded.

LC-MS analysis:

Extracted analytes were separated and analyzed using the Vanq HPLC (ThermoFisher Scientific) coupled to the Sciex7500 triple qu mass spectrometer (SCIEX).

Data processing:

Raw files were processed using Skyline v.23.1. Statistical evaluati data was done via MetaboAnalyst v5.0.

ion and	List of internal standards, concentrations, and re
netabolite	Internal Standard [pmol/µL] CV [%] 0 20 40
tes. vet weight hroughout	Tyrosine 500 Phenylalanine 500 Adenosine 18 Thiamine 19 Citrulline 500 Ornithine 500 Glutamate 500 Leucine 500 Methionine 500 Arginine 500 Valine 500 Sucrose 14
<image/>	Sphingosine (317) 27 Sphingosine (317) 18 SM (18:1) (d9) 41 PE (15:0-18:1)(d7) 7 Cer (d18:1/16:0) (d7) 2 GluCer (d18:1/18:0) 1 Cholesterol (d7) 635 LysoGB3 (d7) 1 MG (18:1) (d7) 5 DG (15:0-18:1) (d7) 17 PA (15:0-18:1) (d7) 11 hemi BMP (14:0) 6 BMP (14:0) 8 LPE (18:1) (d7) 10 PI (15:0-18:1) (d7) 12
scle, brain, ble biopsy ch well of (MeOH). BeatBox at	To determine the robustness of metabolite extractio with low-input, a pool (12 polar metabolites, 21 nor different stable isotope-labeled standards (IS) were samples before homogenization. Overall, the extra resulted in a CV of the extracted metabolites of below a robust and reproducible workflow.
quish Duo Juadrupole	
tion of the	 The BeatBox platform offers a fast, efficient, and A novel high-throughput BeatBox processing work Highly efficient approach for low-input, high-throu Extraction of polar metabolites and non-polar lillarge clinical cohorts.

• Top 10 selected polar metabolites and non-polar lipids indicated biological trends of the tissue origin.



n-polar lipids) of

The top 10 polar metabolites revealed unique regulation levels throughout various tissues. Creatine is upregulated in the PC1 (42.4%) brain, muscle, and cardiac muscle tissues, while ornithine is significantly upregulated in the liver and thiamine upregulated e spiked to the Clustering based on their origin reveals different action efficiency metabolite and lipid level in different organs in the intestine, liver, and brain. In the lipid fraction, glycerophospholipids, like LPC, PC, and PE are upregulated in w 20%, indicating indicating information can be conserved by this cardiac muscle, brain, liver, and muscle tissue. workflow.

KEY TAKEAWAYS

space-saving method for homogenizing various tissue and cell types. kflow for subsequent LC-MS based metabolomic and lipidomic analysis. ughput metabolomics and lipidomics studies.

ipids from limited tissue samples offers an attractive solution for the analysis of

Heatmap of the 10 most abundant analytes



More Information & Contact



Junhua Wang, and Daniel Itzhak at Altos Labs providing the tissue samples and instrumentation 1 this application.

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conflict of Interest Disclosure: K. Limm, S. Wuertenberger, C. Ellis are employed by PreOmics GmbH/PreOmics Inc. B. Ngo, D. Itzhak, J. Wang are employed by Altos Labs

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